

Waterfall Spray zones in Wells Gray Provincial Park: Biodiversity Hotspots and Potential Refugia in a Changing Climate

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Cover: Dawson Falls forms a Niagara-like curtain of water that partly plunges and partly cascades into a broad, shallow canyon. Notably, its spray zone is extended downstream for a distance of nearly 1 km by three smaller cataracts. Photo: Chance Breckenridge

Summary/Blog Post:

Wells Gray Provincial Park is sometimes referred to as “the waterfall park”. One of the major draws for tourism to the park are its waterfalls, most notably Helmcken, Dawson, Moul, and Rainbow Falls, which are visited by tens of thousands annually. But Wells Gray’s waterfalls have considerable significance beyond tourism. Here we focus on their dual function as floristic hotspots and vegetational refugia against climate change – a line of research that has received scant attention to date. We also assessed the degree to which current visitation has degraded waterfall habitats.

The vertical plunge of large waterfalls creates continuous outflow winds that carry mist and aerosolized nutrients tens or, in some cases, hundreds of meters downstream. This mist has a moderating effect on humidity and temperature, cool in summer, and warmer in winter. The net result is a unique set of ecological conditions often suited to the establishment of lichens, mosses, hepatics and plants that are otherwise regionally absent or rare. In short, waterfall spray zones are biodiversity hotspots. In principle, spray zones can also be expected to provide a degree of ecosystem resilience against deepening climate change. While other habitats seem set to become increasingly vulnerable to wildfire, drought, heat and wind, the conditions associated with spray zones are more likely to remain stable through time.

Assessment in 2023 found that floristic richness in the spray zones of Are, Dawson, Moul, and Rainbow falls was significantly higher than observed in habitats upstream. Among all spray zones, 866 species were recorded, almost twice as many species (451) as were recorded among all comparison areas. In total, 536 species of hepatics (49), lichens (304), mosses (126), and 57 vascular plants (57) occurred exclusively within spray zones, compared with 121 species that occurred only outside of spray zones, i.e., 3 hepatics, 49 lichens, 22 mosses, and 47 vascular plants. Thus, the spray zone sites collectively supported more than four times as many exclusive species. Climate monitoring at Are Falls confirmed the moderation of summer climate extremes and the longer duration of potential growth periods for cryptogamic plants.

Introduction.

The primary objective of our study is to call attention to the role of Wells Gray's waterfalls in creating and, through their status as refugia, sustaining nodes of high floristic diversity. Side benefits of our work will include: (1) baseline studies against which future floristic shifts brought about by climate change can be assessed; (2) insights useful in gaining national and international recognition for the values protected in BC's parks, here Wells Gray Park in particular; (3) an interpretive story that deepens our collective appreciation for the Clearwater Valley; and (4) guidance for managers to minimize visitor impacts on sensitive biota.

Prior to the research reported here, no comprehensive studies have been conducted on waterfall spray zone ecosystems in British Columbia. Studies from other parts of the world, however, have highlighted the importance of waterfall spray zones to regional and local ecosystems and biodiversity: Bressard 1972, Kallio 1969, Odland et al. 1991, Zilihona & Nummelin 2001, Quinn et al. 2005.

Waterfall Lichen and Plant Species Inventories (2023).

Lichen, moss, hepatic and plant species were assessed in the spray zone of three waterfalls within Wells Gray Provincial Park - Dawson Falls, Moul Falls, and Rainbow Falls - and one waterfall - Are Falls (Philip Creek) - that occurs just outside its boundaries (Figure 1).

Vegetation assessments at these four waterfalls followed methodologies outlined in the British Columbia Resource Inventory Standards Committee (RISC) Standards (RISC 2024). This methodology employs a "controlled intuitive meander" approach, in which field effort is allocated to all microhabitats present within each waterfall spray zone. Survey routes were therefore not plot- or transect-oriented, but emphasized habitats with potential for maximum species richness, consistent with the goal to compile a comprehensive species list for each waterfall spray zone.

For each of these focal microhabitats, our search effort was determined by a species-area accumulation slope, such that search effort continues only as long as it yields additional species. Once no additional species were located within a defined search area, assessment efforts were shifted to the next selected microhabitat. To compare floristic inventory results of waterfall spray ecosystems against adjacent riparian habitat (e.g. as shown in Fig. 2 for Dawson Falls), each inventory was paired with a second inventory conducted in a roughly equivalent area outside the area of waterfall influence (following the same intuitive search method as described above); where possible, this second inventory was conducted in streamside forests immediately above the waterfall (see Fig. 2).

Ideally, each upstream comparison zone would be contiguous with its corresponding spray zone. Unfortunately, this was not possible in two cases owing to the presence of additional cataracts immediately upstream, each with its own spray zone. Thus, the comparison zone for Moul Falls was positioned at some distance upstream along a low-grade span of the creek, while that for Rainbow Falls had to be situated at some distance downstream. In order to mitigate any residual

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spray zone effects on this later comparison area, and because the comparison area presents only uniformly shrubby fringe habitat, we established a second comparison area along the adjacent shore of Azure Lake.

In total, our final data spreadsheet included 2295 species identifications that enabled a robust comparison of floristic richness within the four spray zones and their corresponding comparison sites. Contributing to this total were 456 specimens made in connection with our study as well as 416 photographs taken and 1,105 voice recordings. The photographs and voice recorder files will be made available upon request; specimens will be available for review once they are accessioned, databased, and filed by UBC Herbarium staff. In order to bolster our findings after the fact, we also included 318 specimens collected in previous years.

In total, we found 987 species of cryptogams and plants in the spray zones and comparison sites, i.e., including 76 hepatics, 476 lichens, 212 mosses, and 223 vascular plants (Fig. 3). Of these, 866 species occurred in waterfall spray zones, versus only 451 species in our comparison sites. Intriguingly, 536 species were recorded exclusively in spray zones, including 49 hepatics, 304 lichens, 126 mosses, and 57 vascular plant versus only 121 species in the comparison sites, i.e., including 3 hepatics, 49 lichens, 22 mosses, and 47 vascular plants. Thus not only did the spray zone sites yield nearly twice as many species as the comparison sites did, they also supported more than four times as many exclusive species.

Each of the spray zones had higher species richness than its corresponding comparison area (Fig. 3). The spray zones of Dawson, Moul, and Rainbow Falls all had more than twice the overall species richness of their corresponding comparison areas. However, vascular plants diverge from this trend compared to bryophytes and lichens: the comparison areas of Are and Dawson Falls had more vascular plants than their corresponding spray zone areas. Removing vascular plants from the species totals, the remaining cryptogam (lichens and bryophytes) floras show especially strong differences: 1.9 (Are), 4.3 (Dawson), 3.5 (Moul), and 3.7 and 2.3 (Rainbow) times higher than in their corresponding comparison areas.

Even more striking is the occurrence of rare or otherwise noteworthy species, which are many times more numerous in the spray zones than in the comparison sites (Table 1); see also Fig. 4. Among the species recorded in the project that are accorded conservation status (Blue or Red) by the British Columbia Conservation Data Centre (BC-CDC 2024), 137 occurred in sprayzones as compared with only 13 species outside the spray zones. Perhaps even more remarkable in this context is existence of 48 spray zone species for which no names could be found. Several of these species may be new to science and, though a few have been observed elsewhere (Björk, unpublished records), they appear to be rare throughout their range in BC. In marked contrast, almost all the lichens, bryophytes and vascular plants observed in the comparison sites outside of waterfall spray zones are common species with broad distributions (with the exception of five).

Environmental Monitoring

Five climate stations were installed in the vicinity of Are Falls during the summer of 2023. Three spray-zone climate stations were installed, one in the immediate spray zone beneath the waterfall (installation in fall 2023), and one in the zone of cool air drift below the waterfall at the lower edge of the spray zone (installation in spring 2023). Data from these climate stations will be contrasted with those from two comparison sites, one immediately adjacent to Are Falls on the upland bench ca. 30 m elevation above the waterfall site, and the second climate station located adjacent to a nearby wetland at Edgewood Blue, providing a comparison site with elevated humidity, but not the cool air drainage and mist associated with the spray zone.

All installations used Hobo H21-002 Microstation Dataloggers (Onset Computer, Cape Cod, MA, USA). Sensors were placed on a mast at 2 m above the ground in canopy openings adjacent to the stream or in the upland forest (see Fig. 5). The one exception was the canopy station in the immediate mist zone at Are Falls, which was installed on the bole of streamside tree due to safety and access constraints at this location.

Temperature and relative humidity (RH) were measured using a Hobo 12-bit Temperature/Relative Humidity Smart Sensor installed in an un aspirated radiation shield. Photosynthetically active radiation (PAR) was recorded using a Hobo Photosynthetic Light (PAR) sensor and incident precipitation measured using a Hobo tipping bucket rain gauge (0.2 mm resolution), while the duration of periods of both incident and occult precipitation (condensation of mist and fog on foliage) measured using Hobo Leaf Wetness Smart Sensors installed at a 45° orientation from vertical. Measurements were taken from all sensors at 5-minute intervals. PAR and precipitation measurements were taken only at the EWB site. We did not install climate stations at the falls where tourism activity is frequent due to concerns about vandalism or tampering in these high-use visitor areas.

Preliminary results from the Are Falls climate stations indicate that the waterfall spray zone and its associated drift of cool moist air downstream has a strong moderating influence on local microclimate compared to surrounding upland forest. Stream-side forests in the extended downstream spray zone (outside of the zone of direct mist deposition - see Fig 2 for relative placement) and adjacent upland forests experienced similar nighttime temperature extremes; however, daytime temperature extremes were strongly ameliorated in the Are falls extended spray zone, with maximum daily temperatures in the upland site often 15 to 20 °C warmer on warm summer days. When combined with often greater humidity found in the streamside forest, this results in an extended duration of potential wetting episodes, as indicated by wetness sensors. This is a critical factor in facilitating the establishment and growth of non-vascular plants. Our climate station installations remain in place and will be monitored through 2024 to further document the extent/significance of spray zone influence.

Our preliminary plans for 2023 research included the examination of water chemistry in the spray zone of study sites. We have deferred this planned work until 2024 due to the greater than anticipated time and resources required to establish climate stations in 2023.

Visitor impact on the floristics of waterfall spray zones

Observation of physical habitats adjacent to Are, Dawson, Moul, and Rainbow Falls show quite different impacts of past human use, correlated with the intensity of visitor use. Are Falls was the least disturbed site, with streamside vegetation and adjacent upland forest showing little human influence. Streamside habitats at Rainbow Falls were also largely undisturbed by human impacts (Fig. 7). Dawson Falls, as a major visitor destination (see cover photograph), showed localized trampling of streamside vegetation, mainly in areas immediately adjacent (both upslope and down to the river shore) to access trails and viewing platforms. This included breakage of canopy twigs and loss of bryophyte and lichen cover in potentially species-rich streamside habitats. At Moul Falls, bryophyte cover is largely absent from areas adjacent to informal trails that lead behind the waterfall, in areas that would otherwise likely have had continuous bryophyte cover (Fig. 8). These latter observations notwithstanding, field work in connection with the present study suggests that human disturbance does not at present threaten species richness within or adjacent to the spray zones of any of the four waterfalls examined, though some species may be threatened if they occur only adjacent to foot-traffic areas.

Wells Gray Park, our study area, falls within the traditional territory of the Simpcw (North Thompson) and Tsq'escenemc (Canim Lake) First Nations. We have discussed our proposal and preliminary research findings with members of Simpcw and have since been in conversation with Mandy Ross, Intermediate Biologist with the Simpcw Resources Group, who is leading discussions on this proposed research with the Simpcw. Our preliminary plans are to host a community workshop at Wells Gray in fall 2024 to share project findings and strategize on meeting future community needs. We anticipate that our findings will prove helpful to coming generations of Indigenous Peoples faced with managing their traditional territory in a time of deepening climate change.

2023 Project budget. The primary project expenditures from BC Parks funds were for species assessments by Curtis Björk and Trevor Goward (Enlivened Consulting, \$13,125.00). The second major project expenditure was for travel and accommodation costs from Prince George to Wells Gray Park (\$1,926.00) (Coxson and Ross). Remaining project expenditures were for replacement probes for the Hobo microclimate stations and related lab supplies (\$2,285.00). UNBC overhead charges of \$927.00 were a final expense. Another \$10,500.00 was provided to Enlivened in 2023 for the waterfall project work from UNBC research funds (BC Real Estate Foundation).

Proposed 2024 Wells-Gray Fieldwork.

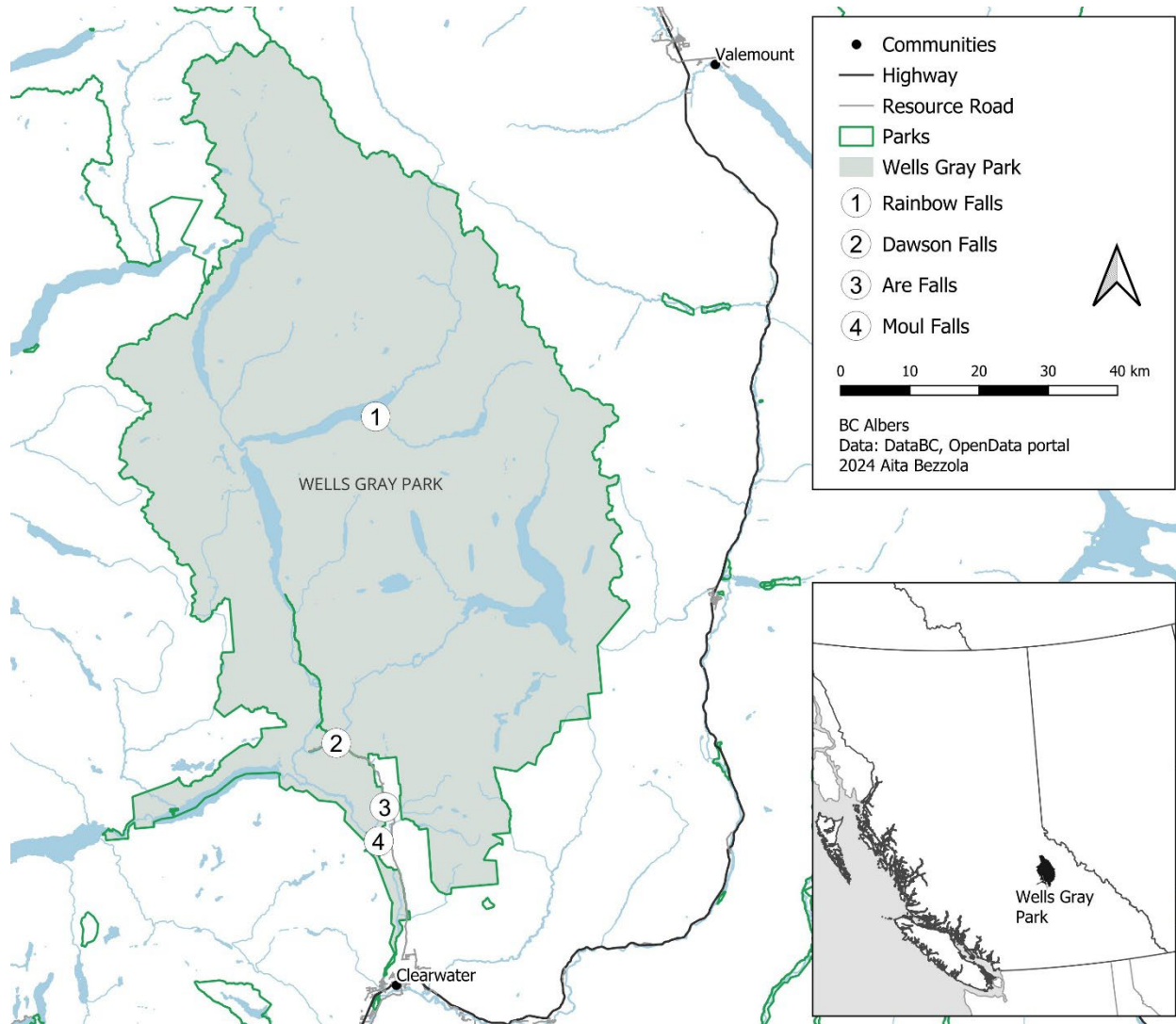
Our proposed research in year two would expand our focus to include two additional heavily visited waterfalls in Wells Gray Park, i.e., Helmcken and Spahats Falls (both major visitor attractions) and Sticta and Triple Decker Falls (both less often visited but with known high biodiversity). We would continue measurements of climate change refugia (microclimate) values at Are falls and would develop an occult precipitation (waterfall mist) collector to assess nutrient contributions in waterfall spray zones.

Literature.

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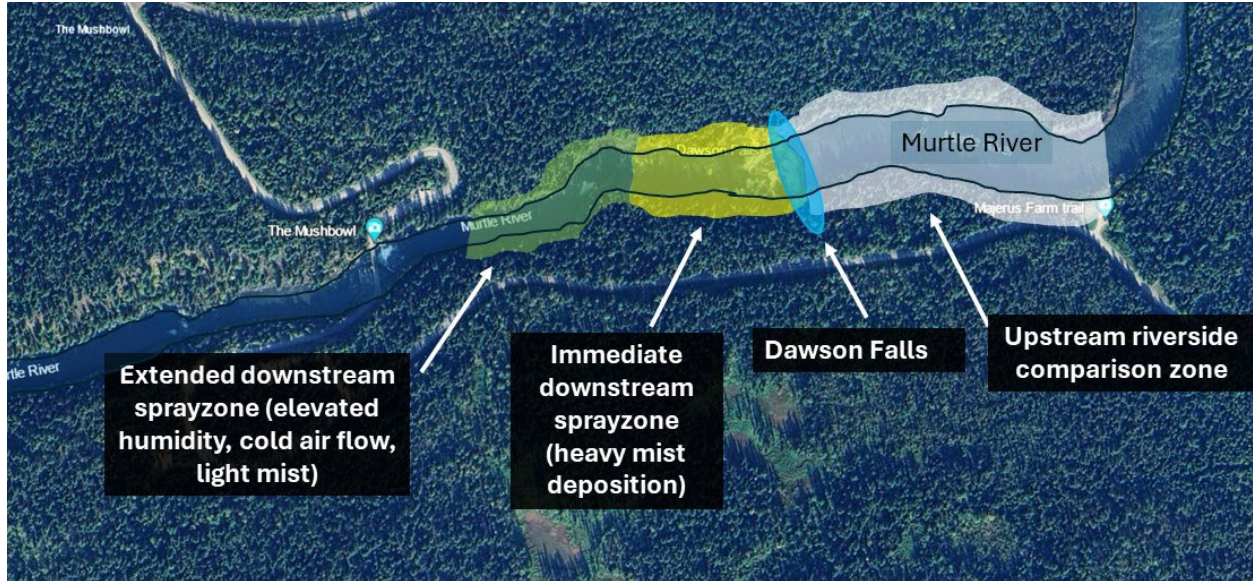
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Figure 1. Map of study area showing location of study sites at Are Falls, Dawson Falls, Moul Falls, and Rainbow Falls within or immediately adjacent to Wells Gray Provincial Park.



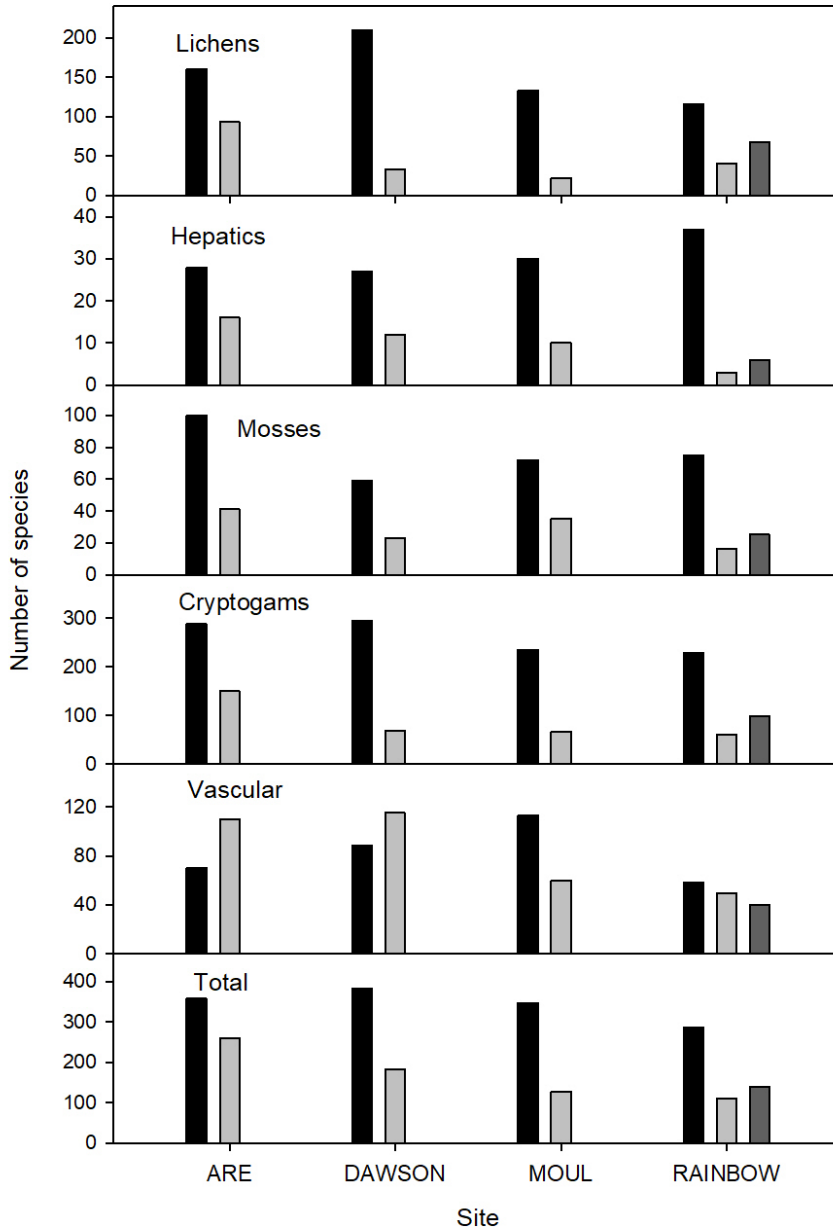
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Figure 2. Potential zones of influence from a waterfall spray zone, illustrated here for Dawson Falls on the Murtle River in Wells Gray Provincial Park. These include the immediate downstream spray zone, characterized by a continued mist deposition on vegetation and exposed surfaces and a strong flow of cool moist air, the zone of extended spray zone influence, with occasional to no direct mist deposition but a strong cold air flow and elevated humidity. Adjacent upstream riverside forests provide a comparison within the landscape for evaluating spray zone influence.



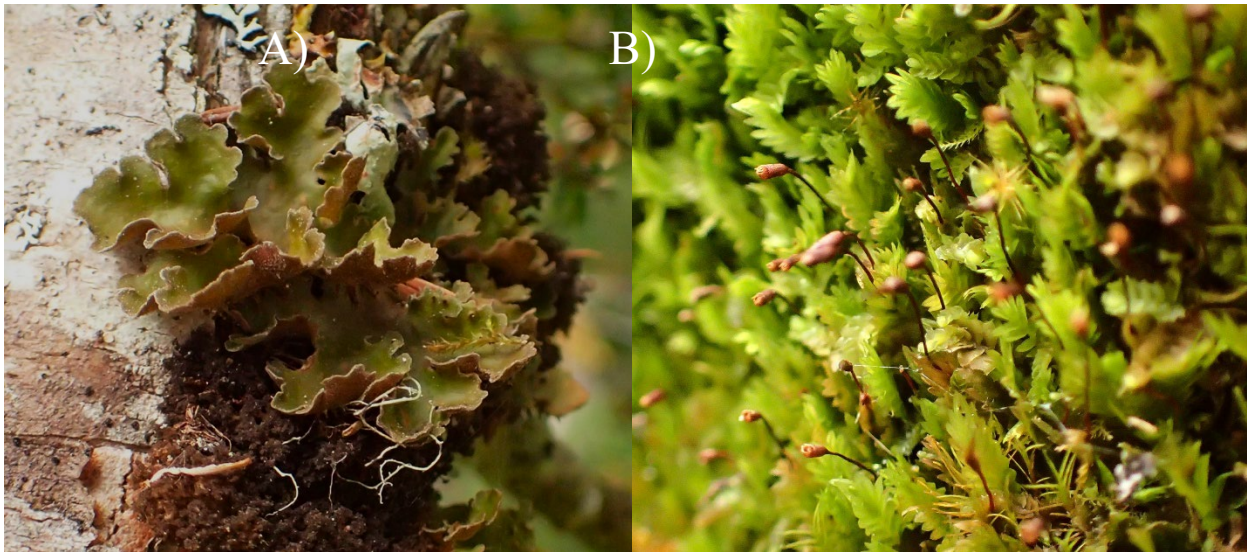
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Figure 3. Species richness by group (hepatics, lichens, mosses, cryptogams, vascular plants, total) for Are, Dawson, Moul, and Rainbow Falls, comparing number of species in the riverside spray zone and in equivalent habitat exposures upstream of the falls. At Rainbow Falls the upstream comparison site was not available due to steep terrain, thus the comparison provided is on a downstream section of Angus Horne Creek, with an additional comparison on the shoreline of Azure Lake.



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Figure 4. Examples of noteworthy species found in the spray zones: A) *Dendrioscicta oroborealis* and B) *Fissidens taxifolius*. *D. oroborealis* is known primarily from the northern Coast Mountains and wettest portion of the northern interior of British Columbia and in the Panhandle of Alaska (COSEWIC 2004, Goward & Björk unpublished data) where it is epiphytic on conifer branches in humid old-growth rainforests (Tønberg and Goward 2001). This is a red-listed species in BC with a provincial status of S1S3 (BC CDC 2024). B) *F. taxifolius* has been reported from coastal BC historically, but its occurrence in the province is not currently accepted. Should any historical records be correctly identified as that species, then it is a new find for interior BC. Otherwise, it is entirely new find as a member of the British Columbia bryoflora.



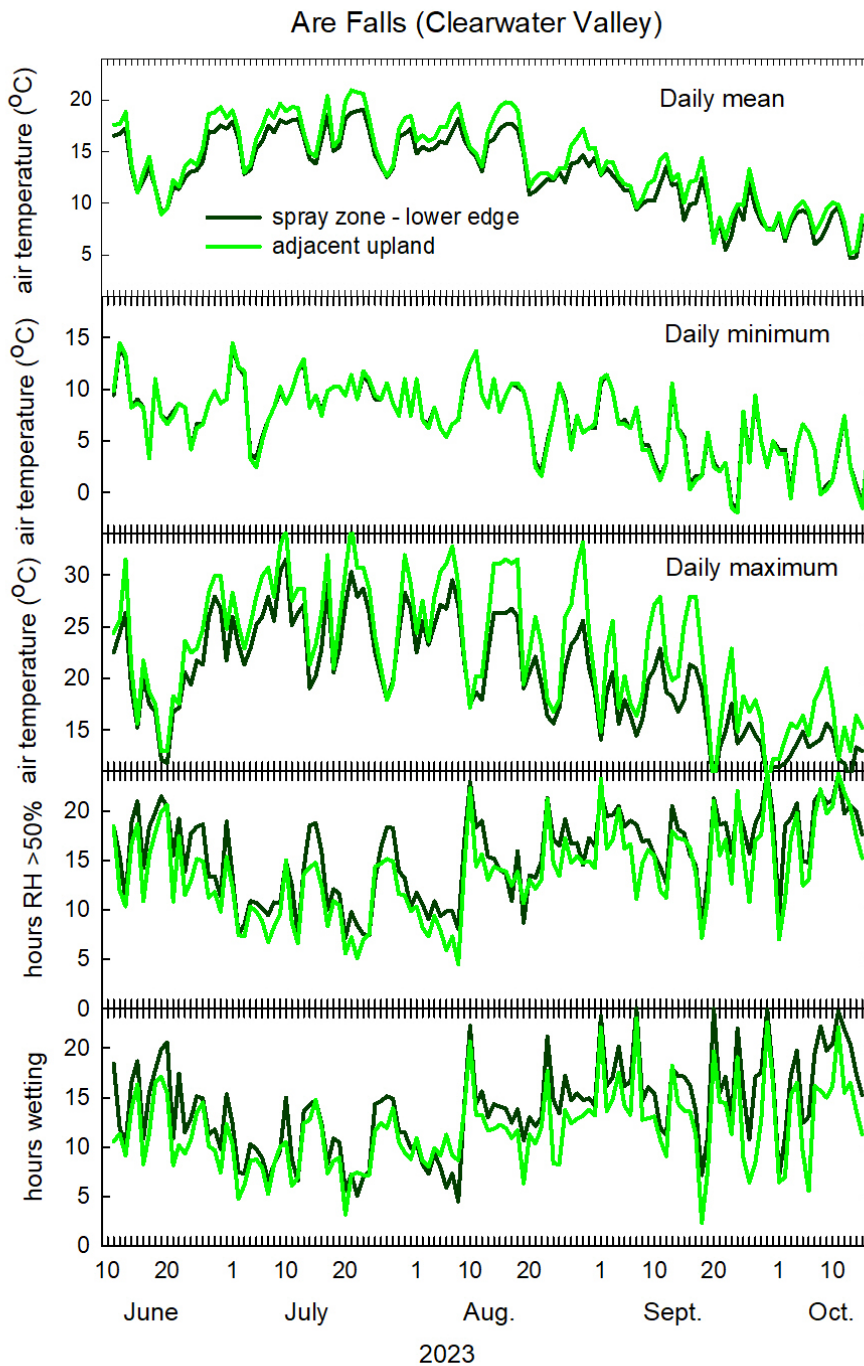
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Figure 5. A) Are Falls has a steep fan-like drop of ca. 7 m over a hard basalt cap, onto more erodible pillow basalt and vesicular basalt. The climate station was placed on a streamside tree adjacent to the creek (outside of image on lower right). B) The lower Are Falls climate station was placed at the outer (lower) edge of the zone of direct spray influence, adjacent to the steep rocky cataract that characterizes downstream flow at Are fall.



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Figure 6. Spray zone microclimate for Are Falls in summer 2023 showing comparisons between the lower edge of the spray zone (black line) and the adjacent upland forest (green line). Comparisons include (from top): daily mean temperatures; daily minimum temperatures, daily maximum temperatures; the daily duration of periods of high humidity; and the daily duration of wetting (when leaf wetness sensors had water sitting on their surface).



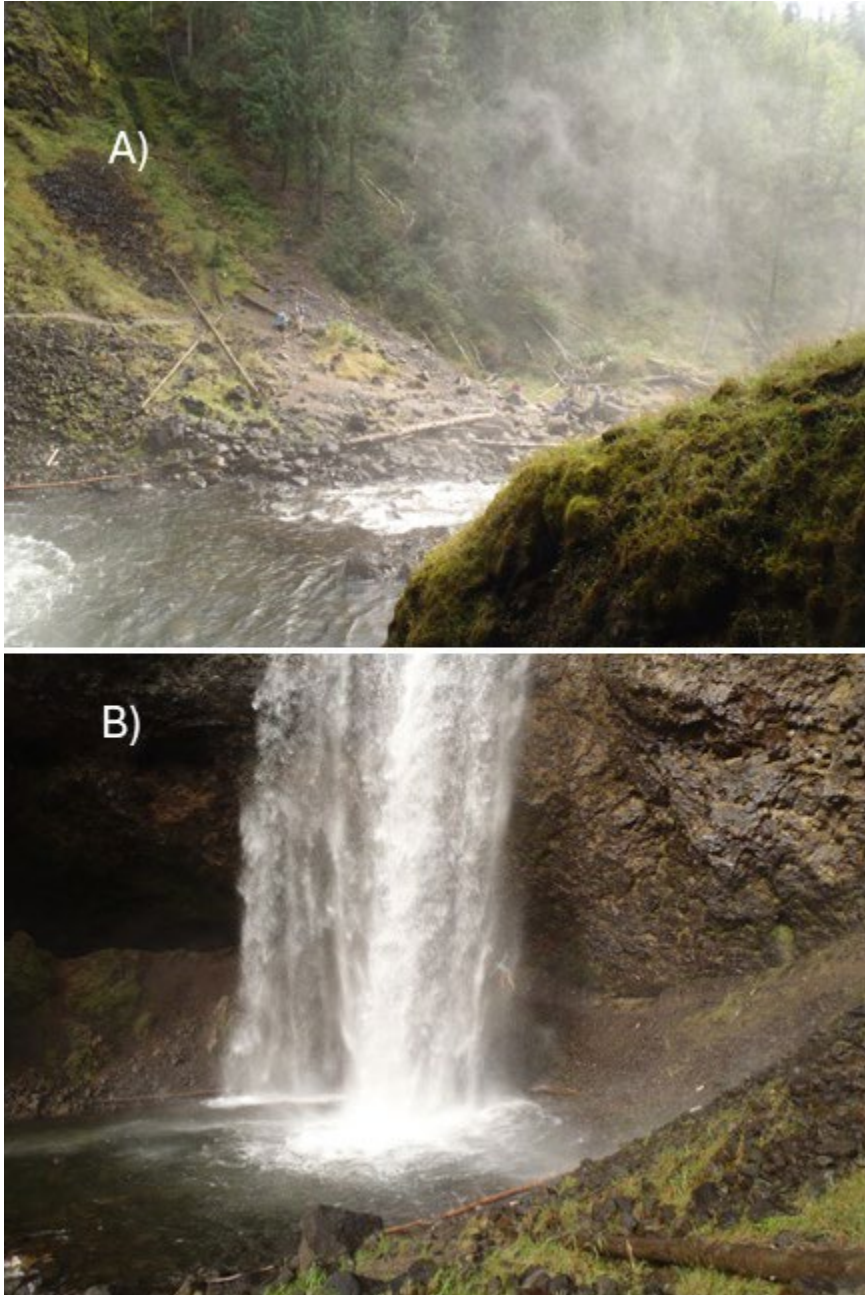
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Figure 7. Waterside habitats at Rainbow Falls are largely undisturbed by human impacts, with viewing access limited to a single overlook at the terminus of a trail.



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Figure 8. Visitor access in areas adjacent to trails used to access the waterfall at Moul Falls has led to the widespread loss of bryophyte cover. A) Trail approaches to Grouse Creek below falls shows extensive areas of bare substrates where vegetation communities have been lost. B) The informal trail behind Moul Falls runs through an area that would otherwise be expected to have widespread bryophyte cover.



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Table 1. Distribution of observed noteworthy species. The first four rows are the spray zone sites; the last five are the corresponding comparison areas outside spray zones. Angus Horne and Azure Lake are the comparison areas corresponding to Rainbow Falls. “CDC rare” is the number of species listed under Blue or Red conservation status by the British Columbia Conservation Data Centre as of January 2024. “Other BC rare” is the number of additional species that are rare in the province. “Rare disjunct” is the number of coastal species that are rare in interior regions. “Rare novelties” is the number of species that may be undescribed species and that are rare in BC.

Site	Total notew.	CDC rare	Other BC rare	Rare disjunct	Rare novelties
Are Falls	19	14	16	2	7
Dawson Falls	53	11	21	3	18
Moul Falls	25	9	6	5	5
Rainbow Falls	59	18	19	7	15
Philip Creek	1	1	0	0	0
Murtle River	0	0	0	0	0
Grouse Creek	9	2	4	2	1
Angus Horne	1	0	0	0	1
Azure Lake	6	2	0	1	3

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Appendix 1. see attached.

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Appendix 1. List of observed species by life form (hepatics, lichens, mosses, vascular plants) in spray and non-spray zones of Are, Dawson, Grouse (Moul), and Rainbow Falls. “Rare coastal disjunct” refers to species that are rare in the interior of British Columbia but more common on the coast. “BC rare species”: BC Conservation Data Centre S-ranks are shown for tracked species. “Are non-spray” is the riparian zone along Philip Creek immediately upstream from Are Falls. “Dawson non-spray” is the riparian zone along the Murtle River immediately upstream from Dawson Falls. “Moul non-spray” is the riparian zone along Grouse Creek in the nearest upstream low-grade span of the creek, separated from Moul Falls by a span of Grouse Creek where numerous smaller cataracts occur. “Angus Horne Creek” is the comparison area for Rainbow Falls, comprising the downstream portion of Angus Horne Creek from the outermost edge of the spray zone to the shore of Azure Lake. “Azure Lake” is the comparison area for Rainbow Falls, comprising the lakeshore habitats from the mouth of Angus Horne Creek to the north end of the Rainbow Falls Campground.

Taxon by Life Form	Rare coastal disjunct	BC rare species	Rare novelties	Note-worthy all	Are spray	Are non-spray	Dawson spray	Dawson non-spray	Moul spray	Moul non-spray	Rain-bow spray	Angus Horne Creek	Azure Lake
<i>Hepatics</i>													
<i>Aneura pinguis</i> (L.) Dumort.							+				+		
<i>Aneura</i> sp. nov.			+	+			+						
<i>Barbilophozia barbata</i> (Schmidel ex Schreb.) Loeske					+	+	+	+					
<i>Barbilophozia hatcheri</i> (A.Evans) Loeske					+				+				
<i>Blasia pusilla</i> L.									+				
<i>Blepharostoma trichophyllum</i> (L.) Dumort.					+	+	+	+	+	+	+		+
<i>Calypogeia integristipula</i> Steph.		S3		+							+		
<i>Cephaloziella divaricata</i> (Sm.) Schiffn.							+						+
<i>Cephaloziella hampeana</i> (Nees) Schiffn. ex Loeske										+			
<i>Cephaloziella turneri</i> (Hook.) Müll. Frib.		S3		+			+						
<i>Chiloscyphus pallescens</i> (Ehrh.) Dumort.						+			+				
<i>Chiloscyphus polyanthos</i> (L.) Corda var. <i>polyanthos</i>								+	+				
<i>Chiloscyphus polyanthos</i> var. <i>rivularis</i> (Schrad.) Debat									+				
<i>Conocephalum conicum</i> (L.) Dumort.							+				+		

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<i>Syzygiella autumnalis</i> (DC.) K.Feldberg, Vána, Hentschel & Heinrichs						+	+				+			
<i>Trilophozia quinqueidentata</i> (Huds.) Bakalin											+			
<i>Tritomaria exsecta</i> (Schmidel) Schiffn. & Loeske		S3				+	+						+	
<i>Tritomaria exsectiformis</i> (Breidl.) Schiffn. ex Loeske		S3				+	+	+	+		+		+	
<i>Tritomaria scitula</i> (Taylor) Jørg.		S2S3				+	+				+		+	
Lichens														
<i>Acarospora fuscata</i> (Nyl.) Th.Fr.						+	+	+						
<i>Acarospora heppii</i> (Nägeli ex Hepp) Nägeli													+	
<i>Acarospora smaragdula</i> (Wahlenb.) A.Massal.						+								
<i>Acarospora</i> unknown species (bullate)						+								
<i>Agonimia tristicula</i> (Nyl.) Zahlbr.						+								
<i>Agonimia tristicula</i> (Nyl.) Zahlbr. aff. (brown spores and pyriform perithecia)				+	+									
<i>Agyrium rufum</i> (Pers.) Fr.								+						
<i>Alectoria sarmentosa</i> (Ach.) Ach.						+	+	+	+	+	+	+	+	+
<i>Amygdalaria panaeola</i> (Ach.) Hertel & Brodo											+			
<i>Arthonia apatetica</i> (A.Massal.) Th.Fr.						+							+	
<i>Arthonia arthonioides</i> (Ach.) A.L.Sm.													+	
<i>Arthonia didyma</i> Körb.		+		+										
<i>Arthonia dispuncta</i> Nyl.		+		+										
<i>Arthonia edgewoodensis</i> Björk ined.								+						
<i>Arthonia lapidicola</i> (Taylor) Branth & Rostr.													+	
<i>Arthonia lignariella</i> Coppins						+								
<i>Arthonia lignariella</i> Coppins aff.				+	+								+	
<i>Arthonia mediella</i> Nyl.						+								
<i>Arthonia muscigena</i> Th.Fr.													+	
<i>Arthonia patellulata</i> Nyl.														+

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<i>Chaenotheca furfuracea</i> (L.) Tibell						+		+					
<i>Chaenotheca gracilentata</i> (Ach.) Mattsson & Middelb.						+		+					
<i>Chaenotheca nitidula</i> Tibell								+					
<i>Chaenotheca stemonea</i> (Ach.) Müll. Arg.						+		+		+		+	
<i>Chaenotheca subroscida</i> (Eitner) Zahlbr.								+				+	
<i>Chaenotheca trichialis</i> (Ach.) Th.Fr.						+							
<i>Chaenotheca xyloxena</i> Nád.v.								+		+			
<i>Chaenothecopsis nana</i> Tibell								+		+			+
<i>Chaenothecopsis nigra</i> Tibell		+			+								+
<i>Chaenothecopsis retinens</i> (Nyl.) Tibell (one ascomatum, sacrificed for microscopy)		+			+								+
<i>Chaenothecopsis tasmanica</i> Tibell								+					+
<i>Chaenothecopsis</i> unknown (growing on <i>Bactrospora</i> apothecia, similar to <i>C. vainioana</i> , but spores smaller)					+	+							+
<i>Chaenothecopsis viridireagens</i> (Nád.v.) A.F.W. Schmidt								+					
<i>Cheiromycina flabelliformis</i> B.Sutton								+					
<i>Chromatochlamys muscorum</i> (Th.Fr.) H.Mayrhofer & Poelt var. <i>muscorum</i>						+							
<i>Chrysothrix candelaris</i> (L.) J.R.Laundon s. lat. (patchy form)										+			
<i>Chrysothrix candelaris</i> (L.) J.R.Laundon s. str.						+	+			+			
<i>Cladonia acuminata</i> (Ach.) Norrl.		+			+	+		+		+		+	
<i>Cladonia arbuscula</i> (Wallr.) Flot.										+			
<i>Cladonia cariosa</i> (Ach.) Spreng. cfr.								+					
<i>Cladonia carneola</i> (Fr.) Fr.						+				+			+
<i>Cladonia cenotea</i> (Ach.) Schaer.						+				+			+
<i>Cladonia chlorophaea</i> (Flörke ex Sommerf.) Spreng. aggr.						+	+	+		+		+	
<i>Cladonia coccifera</i> (L.) Willd.						+							
<i>Cladonia coniocraea</i> (Flörke) Spreng.										+		+	+
<i>Cladonia cornuta</i> (L.) Hoffm. ssp. <i>cornuta</i>						+	+			+			+
<i>Cladonia crispata</i> (Ach.) Flotow subsp. <i>crispata</i>						+		+					

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<i>Cladonia fimbriata</i> (L.) Fr.					+	+		+					+	
<i>Cladonia gracilis</i> subsp. <i>turbinata</i> (Ach.) Ahti					+								+	
<i>Cladonia metacorallifera</i> Asahina					+									
<i>Cladonia multiformis</i> G.Merr.					+								+	
<i>Cladonia novochlorophaea</i> (Simpan) Brodo & Ahti cfr.								+						
<i>Cladonia ochrochlora</i> Flörke								+					+	
<i>Cladonia phyllophora</i> Ehrh. ex Hoffm.					+			+					+	+
<i>Cladonia pleurota</i> (Flörke) Schaer.					+									
<i>Cladonia pocillum</i> (Ach.) O.J.Rich.					+									+
<i>Cladonia pyxidata</i> (L.) Hoffm.					+	+		+					+	
<i>Cladonia rangiferina</i> (L.) Weber ex F.H. Wigg.													+	
<i>Cladonia scabriuscula</i> (Delise) Leight.								+						
<i>Cladonia squamosa</i> (Scop.) Hoffm.					+									
<i>Cladonia stricta</i> (Nyl.) Nyl.					+									
<i>Cladonia subulata</i> (L.) Weber ex F.H.Wigg.					+	+								
<i>Cladonia sulphurina</i> (Michx.) Fr.													+	
<i>Cladonia symphycarpa</i> (Flörke) Fr.					+								+	
<i>Cladonia verruculosa</i> (Vain.) Ahti					+									
<i>Cladonia verticillata</i> (Hoffm.) Schaer.													+	
<i>Cliostomum griffithii</i> (Sm.) Coppins													+	
<i>Cliostomum spribillei</i> Goward & Tønsberg								+					+	
<i>Cliostomum vitellinum</i> Gowan aff.								+	+					
<i>Coenogonium pineti</i> (Ach.) Lücking & Lumbsch								+						
<i>Collema auriforme</i> (With.) Coppins & J.R.Laundon cfr.			+		+			+						
<i>Collema coniophilum</i> Goward			S2, T		+			+					+	
<i>Collema furfuraceum</i> Du Rietz													+	
<i>Collema latzelii</i> Zahlbr.			+		+								+	
<i>Collema occultatum</i> Bagl.								+					+	+
<i>Collema polycarpon</i> Hoffm.								+						

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<i>Collema pseudosaturinum</i> Goward ined.			+	+	+									
<i>Collema subflaccidum</i> Degel.								+					+	
<i>Collema suboceanica</i> Goward ined.			+	+									+	
<i>Collema undulatum</i> var. <i>granulosum</i> Degel. Crust with black apothecia and "halos" of round flat grey areoles (can't collect it)					+									
Cyanolichen, lichinoid? Minute, pulvinate.			+	+				+						
<i>Cyphelium inquinans</i> (Sm.) Trevis.					+					+				+
<i>Dendriscosticta oroborealis</i> (Goward & Tønsberg) Moncada & Lücking chloromorph		S+S3		+									+	
<i>Dermatocarpon bachmannii</i> Anders								+						
<i>Dermatocarpon meiophyllizum</i> Vain.								+						
<i>Dermatocarpon miniatum</i> (L.) W.Mann								+		+				
<i>Diploschistes scruposus</i> (Schreb.) Norman					+					+				
<i>Eopyrenula</i> sp. nov.			+	+				+						
<i>Ephebe hispidula</i> (Ach.) Horw.					+									
<i>Evernia mesomorpha</i> Nyl.										+				
<i>Frutidella pullata</i> (Norman) Schmull					+	+								
<i>Fuscidea gothoburgensis</i> (H.Magn.) V.Wirth & Vězda					+			+						
<i>Fuscopannaria cheiroloba</i> (Müll. Arg.) P.M. Jørg.								+						
<i>Fuscopannaria confusa</i> (P.M. Jørg.) P.M. Jørg.					+			+					+	
<i>Fuscopannaria mediterranea</i> (Tav.) P.M. Jørg.					+			+						
<i>Fuscopannaria praetermissa</i> (Nyl.) P.M. Jørg.					+			+						
<i>Fuscopannaria</i> sp. nov. (isidiate, otherwise similar to <i>F. confusa</i>)			+	+									+	
<i>Gyalecta geioca</i> (Wahlenb. ex Ach.) Ach.		+		+				+						
<i>Gyalecta jenensis</i> (Batsch) Zahlbr.		+		+									+	
<i>Gyalecta russula</i> (Körb. ex Nyl.) Baloch, Lumbsch & Wedin		+		+				+					+	
<i>Haematomma ochroleucum</i> (Neck.) J.R.Laundon					+			+					+	
<i>Hypocenomyce scalaris</i> (Ach. ex Lilj.) M.Choisy										+				
<i>Hypogymnia enteromorpha</i> (Ach.) Nyl.					+	+							+	+

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<i>Mycoblastus sanguinarius</i> (L.) Norman								+		+	+	+	+		
<i>Mycocalicium subtile</i> (Pers.) Szatala												+			
<i>Nephroma arcticum</i> (L.) Torss.								+							
<i>Nephroma bellum</i> (Spreng.) Tuck.												+			
<i>Nephroma helveticum</i> Ach.		S3						+							
<i>Nephroma isidiosum</i> (Nyl.) Gyeln.		S3						+							+
<i>Nephroma parile</i> (Ach.) Ach.												+		+	
<i>Nephroma resupinatum</i> (L.) Ach.														+	+
<i>Nephroma sipeanum</i> Gyeln.								+	+	+	+	+	+		+
<i>Nodobryoria oregana</i> (Tuck. ex Nyl.) Common & Brodo															
<i>Ochrolechia androgyna</i> (Hoffm.) Arnold															+
<i>Ochrolechia gowardii</i> Brodo															
<i>Ochrolechia oregonensis</i> H.Magn.															+
<i>Ochrolechia szatalaensis</i> Verseghy												+			+
<i>Opegrapha zonata</i> Körb.		+										+			
<i>Ophioparma rubricosa</i> (Müll. Arg.) S.Ekman												+	+		
<i>Parmelia hygrophila</i> Goward & Ahti														+	+
<i>Parmelia saxatilis</i> (L.) Ach.													+		
<i>Parmelia sulcata</i> Taylor												+	+	+	+
<i>Parmelia sulymae</i> Goward, Divakar, M.C.Molina & A.Crespo															+
<i>Parmeliella parvula</i> P.M.Jørg.												+			
<i>Parmeliella triptophylla</i> (Ach.) Müll. Arg.													+		+
<i>Parmeliopsis ambigua</i> (Wulfen) Nyl.												+	+	+	+
<i>Parmeliopsis hyperopta</i> (Ach.) Vain.												+	+	+	+
<i>Peltigera aphthosa</i> (L.) Willd.												+		+	
<i>Peltigera britannica</i> (Gyeln.) Holt.-Hartw. & Tønsberg													+		
<i>Peltigera canina</i> (L.) Willd. cfr.													+	+	
<i>Peltigera collina</i> (Ach.) Schrad.													+		+
<i>Peltigera didactyla</i> (With.) J.R.Laundon													+		

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<i>Peltigera elisabethae</i> Gyeln.						+			+				
<i>Peltigera extenuata</i> (Nyl.) Vain.						+	+	+					
<i>Peltigera</i> hairy above veins moderately raised, beige coloured, slow to darken, rhizines not very distinct from each other													+
<i>Peltigera horizontalis</i> (Huds.) Baumg.						+						+	
<i>Peltigera kristinssonii</i> Vitik.										+			
<i>Peltigera lepidophora</i> (Vain.) Bitter						+							
<i>Peltigera leucophlebia</i> (Nyl.) Gyeln.						+				+		+	
<i>Peltigera membranacea</i> (Ach.) Nyl.						+	+	+		+		+	
<i>Peltigera praetextata</i> (Flörke ex Sommerf.) Zopf												+	
<i>Peltigera rufescens</i> (Weiss) Humb.												+	
<i>Peltigera scabrosa</i> Th.Fr.												+	
<i>Peltigera</i> sp. nov. 52692				+	+							+	
<i>Peltigera venosa</i> (L.) Hoffm.						+	+	+					+
<i>Pertusaria alpina</i> Hepp ex Ahles cfr.													+
<i>Pertusaria amara</i> (Ach.) Nyl.										+		+	+
<i>Pertusaria carneopallida</i> (Nyl.) Anzi ex Nyl.												+	
<i>Pertusaria multipuncta</i> (Turner) Nyl.												+	
<i>Pertusaria ophthalmiza</i> (Nyl.) Nyl. aggr.												+	
<i>Pertusaria peculiaris</i> Björk ined.												+	
<i>Pertusaria pupillaris</i> (Nyl.) Th.Fr.						+						+	
<i>Pertusaria sommerfeltii</i> (Sommerf.) Flörke												+	+
<i>Pertusaria</i> sp. 6												+	
<i>Pertusaria subambigens</i> Dibben												+	+
<i>Phaeocalicium betulinum</i> (Nyl.) Tibell													+
<i>Phaeocalicium compressulum</i> (Nyl. ex Vain.) A.F.W. Schmidt												+	
<i>Phaeocalicium populneum</i> (Brond. ex Duby) A.F.W.Schmidt													+
<i>Phaeophyscia decolor</i> (Kashiw.) Essl.						+	+	+					
<i>Phaeophyscia kairamoi</i> (Vain.) Moberg												+	+

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<i>Phaeophyscia sciastra</i> (Ach.) Moberg						+			+					
<i>Phlyctis argena</i> (Ach.) Flot.													+	+
<i>Physcia adscendens</i> H.Olivier									+				+	
<i>Physcia alnophila</i> (Vain.) Loht., Moberg, Myllys & Tehler						+							+	+
<i>Physcia caesia</i> (Hoffm.) Hampe ex Fürnr.									+					
<i>Physcia dubia</i> (Hoffm.) Lettau												+		
<i>Physcia occidentalis</i> Essl. ined.						+								
<i>Physcia phaea</i> (Tuck.) J.W.Thomson						+								
<i>Physconia muscigena</i> (Ach.) Poelt													+	
<i>Physconia perisidiosa</i> (Erichsen) Moberg									+				+	
<i>Physconia</i> sp. nov.														+
<i>Piccolia ochrophora</i> (Nyl.) Hafellner														+
<i>Pilophorus cereolus</i> (Ach.) Th.Fr.									+					
<i>Placopsis fusciculoides</i> D.J.Galloway									+					
<i>Placynthiella dasaea</i> (Stirt.) Tønsberg									+					
<i>Placynthiella icmalea</i> (Ach.) Coppins & P.James														+
<i>Placynthiella uliginosa</i> (Schrad.) Coppins & P.James cfr.														+
<i>Placynthium flabellum</i> (Tuck.) Zahlbr.														+
<i>Placynthium nigrum</i> (Huds.) Gray														+
<i>Placynthium pannariellum</i> (Nyl.) H.Magn.														+
<i>Platismatia glauca</i> (L.) W.L.Culb. & C.F.Culb.									+				+	+
<i>Platismatia norvegica</i> (Lynge) W.L.Culb. & C.F.Culb.														+
<i>Polyblastia</i> sp.														+
<i>Polychidium muscicola</i> (Sw.) Gray														+
<i>Porpidia cinereoatra</i> (Ach.) Hertel & Knoph														+
<i>Porpidia contraponenda</i> (Arnold) Knoph & Hertel														+
<i>Porpidia crustulata</i> (Ach.) Hertel & Knoph														+
<i>Porpidia melinodes</i> (Körb.) Gowan & Ahti														+
<i>Porpidia ochrolemma</i> (Vain.) Brodo & R.Sant.														+

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<i>Brachythecium acutum</i> (Mitt.) Sull. ex Austin						+	+							
<i>Brachythecium asperrimum</i> (Mitt. ex Müll.Hal.) Sull.														
<i>Brachythecium frigidum</i> (Müll.Hal.) Besch.														
<i>Brachythecium rivulare</i> Schimp.														
<i>Brachythecium rotaezanum</i> De Not.														
<i>Brachythecium salebrosum</i> (Hoffm. ex F.Weber & D.Mohr) Schimp.														
<i>Brideliella wahlenbergii</i> (Brid.) Fedosov, M.Stech & Ignatov														
<i>Bryum argenteum</i> Hedw.														
<i>Bryum lanatum</i> (P.Beauv.) Brid.														
<i>Bucklandiella heterosticha</i> (Hedw.) Bedn.-Ochyra & Ochyra														
<i>Bucklandiella microcarpa</i> (Hedw.) Bedn.-Ochyra & Ochyra														
<i>Calliergon cordifolium</i> (Hedw.) Kindb.														
<i>Campylium chrysophyllum</i> (Brid.) Lange														
<i>Campylium protensum</i> (Brid.) Kindb.														
<i>Campylium stellatum</i> (Hedw.) C.E.O.Jensen														
<i>Campylopus atrovirens</i> De Not.														
<i>Campylopus flexuosus</i> (Hedw.) Brid.														
<i>Ceratodon purpureus</i> (Hedw.) Brid.														
<i>Chionoloma tenuirostre</i> (Hook. & Taylor) Lindb.														
<i>Claopodium bolanderi</i> Best														
<i>Climacium dendroides</i> (Hedw.) F.Weber & D.Mohr														
<i>Codriophorus acicularis</i> (Hedw.) P.Beauv.														
<i>Codriophorus mollis</i> (Cardot) Bedn.-Ochyra & Ochyra														
<i>Conardia compacta</i> (Müll. Hal.) H. Rob.														
<i>Cratoneuron filicinum</i> (Hedw.) Spruce														
<i>Ctenidium</i> sp. +														
<i>Ctenidium</i> sp. 2														
<i>Dichodontium pellucidum</i> (Hedw.) Schimp.														

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<i>Orthotrichum pallens</i> Bruch ex Brid.		+		+	+									
<i>Palustriella falcata</i> (Brid.) Hedenäs								+					+	
<i>Philonotis fontana</i> (Hedw.) Brid.								+				+	+	
<i>Philonotis marchica</i> (Hedw.) Brid.		S2S3		+	+			+						
<i>Philonotis tomentella</i> Molendo								+					+	
<i>Plagiopus oederianus</i> (Sw.) H.A.Crum & L.E.Anderson								+		+			+	
<i>Plagiothecium cavifolium</i> (Brid.) Z.Iwats.												+		
<i>Platydictya confervoides</i> (Brid.) H.A.Crum cfr.		+		+	+									
<i>Platydictya jungermannioides</i> (Brid.) H.A.Crum					+	+				+				
<i>Platydictya minutissima</i> (Sull. & Lesq.) H.A.Crum cfr. (but leaves falcate)		+		+				+						
<i>Platyhypnum alpestre</i> (Hedw.) Ochyra		S3		+				+					+	
<i>Platyhypnum alpinum</i> (Lindb.) Loeske		S3		+								+	+	
<i>Platyhypnum duriusculum</i> (De Not.) Ochyra								+				+		
<i>Platyhypnum norvegicum</i> (Schimp.) Ochyra		S2		+									+	
<i>Platyhypnum smithii</i> (Sw.) Ochyra								+						
<i>Pleurozium schreberi</i> (Willd. ex Brid.) Mitt.					+	+	+	+	+	+	+		+	+
<i>Pogonatum dentatum</i> (Menzies ex Brid.) Brid.					+									
<i>Pogonatum urnigerum</i> (Hedw.) P.Beauv.						+							+	
<i>Pohlia andalusica</i> (Höhn.) Broth.		S2		+						+				
<i>Pohlia annotina</i> (Hedw.) Lindb.					+	+				+				
<i>Pohlia cruda</i> (Hedw.) Lindb.					+	+	+			+	+			
<i>Pohlia drummondii</i> (Müll.Hal.) A.L.Andrews						+								
<i>Pohlia longibracteata</i> Broth.	+			+						+	+			
<i>Pohlia nutans</i> (Hedw.) Lindb.					+	+				+				
<i>Pohlia proligera</i> (Kindb. ex Breidl.) Lindb. ex Arnell						+								
<i>Polytrichum hyperboreum</i> R.Br.		S+S3		+	+			+						
<i>Polytrichum juniperinum</i> Hedw.										+			+	
<i>Pseudisothecium stoloniferum</i> (Brid.) Grout													+	+

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<i>Schistidium papillosum</i> Culm.					+	+		+	+	+			
<i>Schistidium pulchrum</i> H.H.Blom	S3			+	+								
<i>Schistidium rivulare</i> (Brid.) Podp.									+				
<i>Schistidium subjulaceum</i> H.H.Blom						+							
<i>Schistidium trichodon</i> (Brid.) Poelt	S3			+					+				
<i>Schistidium</i> unknown species (tiny capsules, red operculum)			+	+	+				+				
<i>Sciurohypnum latifolium</i> (Kindb.) Ignatov & Huttunen								+					
<i>Sciurohypnum plumosum</i> (Hedw.) Ignatov & Huttunen						+				+			
<i>Scleropodium obtusifolium</i> (Mitt.) Kindb.								+					
<i>Scouleria aquatica</i> Hook.					+	+	+	+	+	+		+	
<i>Sphagnum capillifolium</i> (Ehrh.) Hedw.												+	
<i>Sphagnum girgensohnii</i> Russow												+	
<i>Sphagnum warnstorffii</i> Russow												+	
<i>Streblotrichum convolutum</i> (Hedw.) P.Beauv								+					
<i>Syntrichia norvegica</i> F.Weber aff.			+	+	+								
<i>Syntrichia ruralis</i> (Hedw.) F.Weber & D.Mohr						+		+					
<i>Tetraphis geniculata</i> Girg. ex Milde								+		+			
<i>Tetraphis pellucida</i> Hedw.								+					
<i>Thamnobryum neckeroides</i> (Hook.) E.Lawton						+		+		+			
<i>Thuidium recognitum</i> (Hedw.) Lindb.						+				+		+	
<i>Timmia austriaca</i> Hedw.						+		+					
<i>Timmia megapolitana</i> Hedw. ssp. <i>megapolitana</i>						+							
<i>Tortella fragilis</i> (Hook. & Wilson) Limpr.										+			
<i>Tortella nitida</i> var. <i>fragilifolia</i> (Jur.) Köckinger & Hedenäs												+	
<i>Tortella tortuosa</i> (Schrad. ex Hedw.) Limpr.						+		+		+		+	+
<i>Tortula mucronifolia</i> Schwägr.						+							
<i>Trematodon</i> sp.?												+	
<i>Trichostomum tenuirostre</i> (Hook. & Taylor) Lindb.						+		+				+	
<i>Trochophyllohypnum circinale</i> (Hook.) Jan Kučera & Ignatov						+				+		+	+

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<i>Vinealobryum nicholsonii</i> (Crum) R.H.Zander			S+			+	+							
Vascular Plants														
<i>Abies lasiocarpa</i> subsp. <i>bifolia</i> (A.Murray) Silba						+	+	+	+		+	+	+	+
<i>Acer glabrum</i> var. <i>douglasii</i> (Hook.) Dippel							+	+	+	+				
<i>Achillea</i> sp.						+					+			
<i>Agrostis capillaris</i> L.							+							
<i>Agrostis exarata</i> Trin.							+							
<i>Agrostis gigantea</i> Roth						+	+	+		+	+			
<i>Agrostis scabra</i> Willd.						+	+	+	+	+	+	+		
<i>Alnus alnobetula</i> var. <i>fruticosa</i> (Rupr.) H.Winkl.						+	+		+		+	+	+	+
<i>Alnus incana</i> (L.) Moench						+	+	+	+	+	+	+	+	+
<i>Amelanchier florida</i> Lindl.									+	+				
<i>Anaphalis margaritacea</i> (L.) Benth. & Hook.f.							+	+		+		+		
<i>Anemone parviflora</i> Michx.												+		
<i>Angelica genuflexa</i> Nutt.						+	+			+		+		
<i>Antennaria howellii</i> Greene										+				
<i>Antennaria rosea</i> Greene									+					
<i>Apocynum androsaemifolium</i> L.							+							
<i>Aquilegia formosa</i> Fisch. ex DC.							+		+	+			+	
<i>Arabidopsis lyrata</i> ssp. <i>kamchatica</i> (Fisch. ex DC.) O'Kane & Al-Shehbaz						+	+			+				
<i>Aralia nudicaulis</i> L.							+	+	+					
<i>Arctostaphylos uva-ursi</i> (L.) Spreng.									+	+				
<i>Arnica amplexicaulis</i> Nutt. var. <i>amplexicaulis</i>								+	+			+		
<i>Asplenium viride</i> Huds.												+		
<i>Athyrium filix-femina</i> (L.) Roth						+	+	+	+	+	+	+	+	+
<i>Berberis aquifolium</i> var. <i>nutkana</i> (DC.) Marroq. & Laferr.						+	+	+	+	+	+			
<i>Betula papyrifera</i> Marshall						+	+	+	+	+	+	+	+	+

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<i>Calamagrostis canadensis</i> (Michx.) P.Beauv.						+	+	+	+	+	+	+	+	+
<i>Calamagrostis rubescens</i> Buckley										+				
<i>Canadanthus modestus</i> (Lindl.) G.L.Nesom						+	+	+	+	+	+	+	+	+
<i>Cardamine oligosperma</i> Nutt.							+			+				
<i>Cardamine pensylvanica</i> Muhl. ex Willd.						+	+			+				
<i>Carex arcta</i> Boott									+					
<i>Carex brunnescens</i> (Pers.) Poir.								+						
<i>Carex capillaris</i> L.												+		
<i>Carex deflexa</i> Hornem.										+				
<i>Carex disperma</i> Dewey									+					
<i>Carex kelloggii</i> W.Boott								+	+	+				+
<i>Carex leptalea</i> Wahlenb.									+					
<i>Carex mertensii</i> J.D.Prescott ex Bong.								+				+		
<i>Carex pedunculata</i> Muhl. ex Willd.						+	+							
<i>Carex rossii</i> Boott									+					
<i>Carex sitchensis</i> J.D.Prescott ex Bong.									+					+
<i>Carex vaginata</i> Tausch									+			+		
<i>Carex viridula</i> Michx.									+				+	+
<i>Castilleja purpurascens</i> Greenm.									+					
<i>Cerastium fontanum</i> Baumg.								+		+				
<i>Chimaphila umbellata</i> (L.) Nutt.										+		+		
<i>Cicuta virosa</i> L.						+	+							
<i>Cinna latifolia</i> (Trevir. ex Göpp.) Griseb.						+	+	+	+	+	+			
<i>Circaea alpina</i> L. ssp. <i>alpina</i>						+	+	+	+	+	+	+	+	
<i>Cirsium vulgare</i> (Savi) Ten.										+				
<i>Clintonia uniflora</i> (Menzies ex Schult. & Schult. f.) Kunth								+	+	+	+	+		
<i>Conyza canadensis</i> (L.) Cronq.										+				
<i>Cornus stolonifera</i> Michx.						+	+	+	+	+	+	+	+	+
<i>Cornus unalaschkensis</i> Ledeb.								+	+	+		+	+	

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<i>Galium triflorum</i> Michx.						+	+	+	+	+	+			
<i>Geum macrophyllum</i> Willd.						+	+	+	+	+	+			
<i>Geum perincisum</i> Rydb.											+			
<i>Glyceria borealis</i> (Nash) Batch.												+		
<i>Glyceria elata</i> (Nash) M.E.Jones						+	+		+	+				
<i>Glyceria striata</i> (Lam.) Hitchc.							+	+	+	+			+	
<i>Goodyera oblongifolia</i> Raf.								+						
<i>Goodyera repens</i> (L.) R.Br.								+						
<i>Gymnocarpium disjunctum</i> (Rupr.) Ching						+		+	+	+		+	+	
<i>Gymnocarpium dryopteris</i> (L.) Newman							+	+				+		
<i>Heracleum lanatum</i> Michx.										+				
<i>Heuchera glabra</i> Willd. ex Roem. & Schult.						+	+	+	+	+		+		
<i>Hieracium albiflorum</i> Hook.						+		+		+				
<i>Hieracium caespitosum</i> Dumort.							+							
<i>Hieracium piloselloides</i> Vill.							+						+	
<i>Hierochloe odorata</i> (L.) P.Beauv.								+						
<i>Hippuris vulgaris</i> L.									+					
<i>Impatiens aurella</i> Rydb.										+				
<i>Juncus alpinoarticulatus</i> Vill.													+	
<i>Juncus ensifolius</i> Wikstr.							+	+	+				+	
<i>Juncus filiformis</i> L.								+	+					
<i>Juncus mertensianus</i> Bong.													+	+
<i>Juncus tenuis</i> Willd.							+							
<i>Lathyrus ochroleucus</i> Hook.										+				
<i>Leucanthemum vulgare</i> (Vaill.) Lam.							+			+				
<i>Lilium columbianum</i> Leichtlin								+						
<i>Linnaea borealis</i> var. <i>americana</i> (J.Forbes) Hultén						+	+	+	+	+	+	+	+	+
<i>Lonicera involucrata</i> (Richardson) Banks ex Spreng.						+	+	+	+	+	+			
<i>Lonicera tatarica</i> L.						+								

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<i>Symphyotrichum ciliolatum</i> (Lindl.) Á. Löve & D. Löve						+	+		+	+	+			
<i>Symphyotrichum eatonii</i> (A.Gray) G.L.Nesom														+
<i>Symphyotrichum foliaceum</i> (Lindl. ex DC.) G.L.Nesom								+					+	
<i>Symphyotrichum maccallae</i> (Rydb.) comb. ined.									+					
<i>Taraxacum</i> sp. (exotic)						+			+	+				
<i>Taxus brevifolia</i> var. <i>reptaneta</i> Spjut													+	
<i>Thuja plicata</i> Donn ex. D.Don						+	+	+	+	+	+	+	+	+
<i>Tiarella trifoliata</i> var. <i>unifoliata</i> (Hook.) Kurtz						+	+	+	+	+	+	+	+	+
<i>Torreyochloa pauciflora</i> (J.Presl) G.L.Church														
<i>Trifolium aureum</i> Pollich										+				
<i>Trifolium hybridum</i> L.														
<i>Tsuga heterophylla</i> (Raf.) Sarg.						+	+	+	+	+	+	+	+	+
<i>Vaccinium cespitosum</i> Michx.										+				
<i>Vaccinium membranaceum</i> Douglas ex Torr.								+	+	+	+		+	+
<i>Vaccinium myrtilloides</i> Michx.										+				
<i>Vaccinium ovalifolium</i> Sm.													+	+
<i>rbascum thapsus</i> L.										+				
<i>Veronica americana</i> Schwein. ex Benth.						+	+	+	+	+	+			
<i>Veronica serpyllifolia</i> L.														
<i>Viburnum edule</i> Raf.								+	+	+	+		+	
<i>Viola adunca</i> Sm.						+				+				
<i>Viola epipsila</i> Ledeb.														+
<i>Viola incognita</i> Brainerd cfr.						+								
<i>Viola palustris</i> L.														
<i>Woodsia scopulina</i> D.C.Eaton						+							+	